

## Master Thesis Marine Biology

### *Composition and distribution of the macrozoobenthic communities on the shelf off Angola*

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### Zusammenfassung

Diese Arbeit beschäftigte sich mit der Zusammensetzung und der Verteilung der makrozoobenthischen Gemeinschaften an 9 Stationen entlang eines latitudinalen Gradienten (8° bis 17°S) in Wassertiefen von 28 m bis 102 m auf dem Schelf vor Angola. Die Temperatur variierte zwischen 14,5°C im Süden und 17,5°C im Norden. Das Sediment zeichnete sich durch unterschiedliche Beschaffenheit aus.

Die Arbeit stellt nun erstmalig eine Zusammenstellung der benthischen Fauna vor Angola an ausgewählten Stationen dar. Es wurden insgesamt 36 Gefäßproben untersucht. Alle Organismen wurden auf dem möglichst niedrigsten taxonomischen Niveau bestimmt. Von den insgesamt 343 ermittelten Taxa weisen die Polychaeten die größte Dominanz mit ca. 40% auf, gefolgt von den Crustaceen (35%), Mollusken (19%), „Andere“ (6%) sowie den Echinodermaten (1%). Die höchste Diversität mit 125 Taxa wurde bei Station Be71 (9°S) ermittelt. Ein weiterer Peak befindet sich bei Station BM5 (13°S) mit 124 Taxa, wobei die geringste Diversität von 42 Taxa bei Station Ku4 gefunden wurde. Die größte Abundanz weist Station Na5 (15°S) mit 38.332 Individuen/m<sup>2</sup> auf, im Vergleich dazu Station Ku4 (17°S) mit der geringsten Abundanz von 1.188 Individuen/m<sup>2</sup>. Die Biomasse variiert zwischen 10,06 g/m<sup>2</sup> (Station SU4, 10°S) und 216,85 g/m<sup>2</sup> (Station Ku4), und wurde im Süden vorrangig von den Mollusken bestimmt. Die Schlüsselarten sind im Süden an der Mündung des Kunene-Flusses *Nuculana bicuspidata*, *Nassarius vinctus* sowie der Polychaet *Cossura coasta*, während in den nördlicheren Regionen die Polychaeten *Diopatra neapolitana*, *Owenia* sp. sowie *Prionospio* sp. sehr abundant auftraten.

Hinsichtlich der Gesamtbetrachtung zeigt sich kein eindeutiger Trend entlang des latitudinalen Gradienten auf dem Schelf vor Angola. Die südlichen Stationen grenzen sich aufgrund des geringen Sauerstoffgehalts von <1 ml/l mit geringer Diversität und Abundanz sowie hoher Biomasse von den übrigen Stationen ab. Jedoch weisen die Werte für die nördlicheren Stationen signifikante Schwankungen auf, die möglicherweise aus der unterschiedlichen Sedimentbeschaffenheit resultieren könnte. Die höchste Diversität wurde bei Stationen mit großer Korngröße ermittelt, wobei steinige Böden eine geringe Diversität aufwiesen. Die größte Abundanz wurde an einer Station bestimmt, die von einer großen Menge an Diatomeen und Schill dominiert wurde, welche als Nahrungsquelle dient sowie Schutz für benthische Organismen bietet.

## Summary

This thesis dealt with the composition and distribution of the macrozoobenthic communities at 9 stations in 28 m to 102 m depth along a latitudinal gradient (8° to 17°S) on the shelf off Angola. The temperature varied between 14,5°C in the south and 17,5°C in the north. The sediment revealed different textures.

This study is the first that presents a composition of the benthic fauna at several selected locations off Angola. A total of 36 samples were analysed. All organisms were determined on the lowest taxonomic level, whenever possible. The 343 taxa, that were encountered, exhibit the dominance of the polychaetes with about 40%, followed by the crustaceans (35%), mollusks (19%), other (6%) and echinoderms (1%).

The highest diversity was detected at station Be71 (9°S) with 125 taxa in total. Another peak can be found at station BM5 (13°S) with 124 taxa, whereupon the lowest diversity of 42 taxa was determined at station Ku4 (17°S). The highest abundance could be shown at station Na5 (15°S) with 38.332 individuals/m<sup>2</sup>, contrary to that is station Ku4 with the lowest abundance of 1.188 individuals/m<sup>2</sup>. The biomass varies between 10,06 g/m<sup>2</sup> at station SU4 (10°S) and 216,85 g/m<sup>2</sup> at station Ku4, and was mainly influenced by the mollusks in the south. The key species in the south of Angola at the mouth of the Cunene River are *Nuculana bicuspidata* and *Nassarius vinctus*, as well as the polychaete *Cossura coasta*, while the polychaetes *Diopatra neapolitana*, *Owenia* sp. and *Prionospio* sp. occur frequently in the northern regions.

In regard to the overall view, it can be concluded that there is no clear trend along the latitudinal gradient on the shelf off Angola. The southern stations delimit from all other observed stations due to their less oxygen content of <1 ml/l which reasons in a minor diversity, abundance as well as high biomass. However, the values of the northern stations vary significantly that possibly could have resulted from the different sediment textures since the highest diversity was measured at stations of large grain sizes, whereupon rocky bottom exhibited low diversity. The highest abundance was detected at a station that was dominated by diatoms and shell detritus offering protection and serving as nutritional basis for benthic organisms.

# 1 Introduction

## 1.1 State of research

Marine sediments form one of the largest habitats on earth by covering more than 80 % of the ocean floor. In spite of high size variations of the benthos, the benthic biomass is dominated by the macrofaunal invertebrates (>0,5 mm), including many species of polychaetes, crustaceans, mollusks and echinoderms (Lenihan & Micheli 2001).

It is a general rule that the species richness of many animals and plants in terrestrial systems declines from the tropics to the poles. It has long been hypothesised that a similar trend is also present in the sea (Ellingsen & Gray 2002).

In fact, this presumption belongs to one of 3 generally accepted gradients of diversity in the sea, that has been summarized by Levinton (1995) as `The best known diversity gradient is an increase of species diversity from high to low latitudes in continental shelf benthos, in the plankton in continental shelf regions and in the open ocean`.

However, studies on marine fauna revealed different results over the past 2 to 3 decades (Renaud et al. 2009).

Up to now, macrozoobenthic communities on shelves have been already studied worldwide, whereupon research at the Angolan coast has only focused on fisheries, zooplankton and meiozoobenthic communities by i.e. Longhurst (1959), Strømme & Sætersdal (1991), Soltwedel & Thiel 1995 and Soltwedel (1997).

### 1.2 Shelf off Angola

Angola with its approximately 1.600 km long coastline is located in the southwest of Africa at the Atlantic ocean in the southern hemisphere. The shelf, describing the shallow, nearshore seabed up to 200 meters below the sea level, is influenced by several abiotic factors such as temperature, oxygen, salinity, sediment and ocean currents. An overview of these features is given in the next paragraphs.

#### 1.2.1 Temperature, salinity and oxygen

Angola is characterised by a temperate climate in the south and a tropic climate in the north. During the summer from January to April, there is an increased rainfall and runoff from the Congo River in the northern boundary of Angola. The upper water layer consists of Equatorial Water that is characterised by low salinity and high temperature (Wauthy 1977). The oxygen level is usually above 2 ml/l in a depth of about 100 m, decreasing to slightly over 1 ml/l at the edge of the shelf. Surface temperatures of the northern part (to Benguela) are usually 27 to 28°C, whereupon bottom temperatures of 20°C are reached to about 50 m depth. By contrast, the temperature near the bottom in the southern regions is always lower than 20°C.

During winter, a northward flowing coastal current develops in consequence of the strengthened southeast trade winds, with upwelling occurring all along the coast. This phenomenon appears to be well developed from the North to the centre of the Angolan coast. Surface temperatures of the northern region decreases (20 to 22°C). In the southern region where upwelling is at its peak, surface temperatures near the coast decreases to 15°C. At about 50 m depth, oxygen values <2 ml/l are reached, whereas values below 1 ml/l are found at 100 m depth (Bianchi, 1992).

According to Lass *et al.* (2000), Angola Current water usually has a temperature greater than 24°C and a salinity of more than 36,4 psu in the upper mixed layer. Traveling southwards, the temperature of the water mass gradually declines and becomes less saline. During winter and spring, the Angola Current water with temperatures between 27°C and 30°C retreats to the northwest replaced by slightly cooler waters with temperatures between 20

and 26°C (Meeuwis & Lutjeharms 1990). The isotherms run more or less parallel to the coast (Strømme & Sætersdal 1991) (Figure 1).

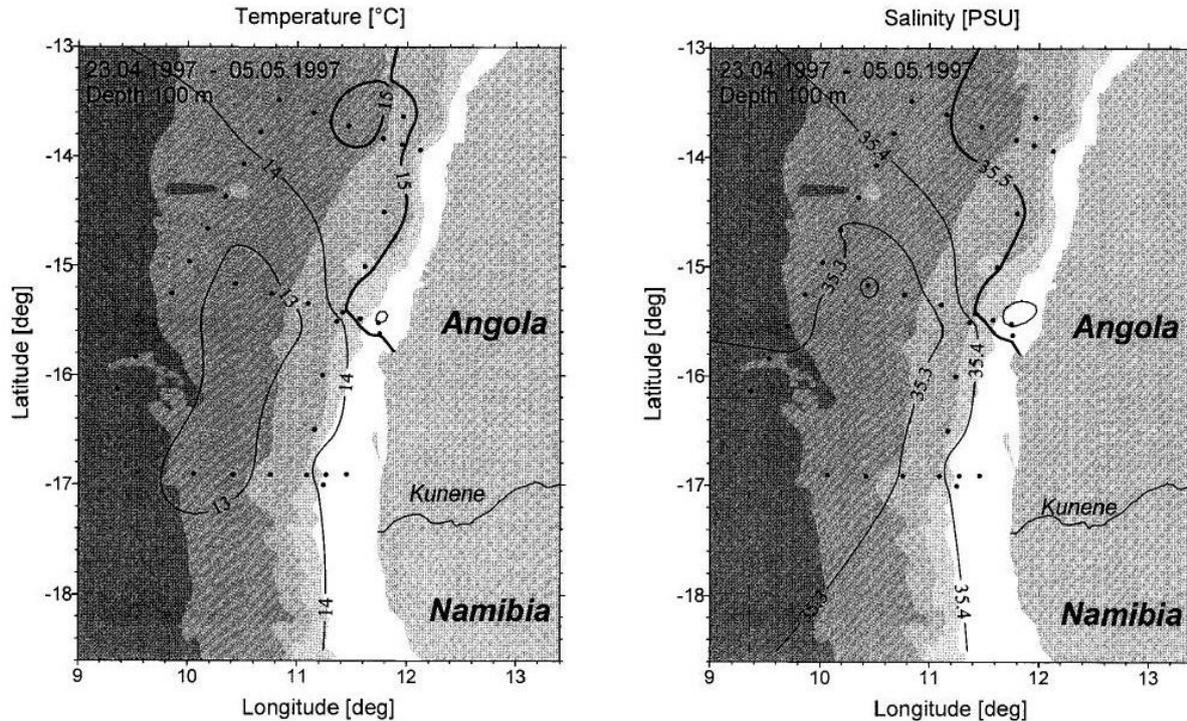


Fig. 1: Horizontal distribution of temperature (left) and salinity (right) in 100 m depth (Lass *et al.* 2000).

### 1.2.2 Bottom topography and structure

There are different bottom structures (Figure 2) and sediment types (Figure 3) along the coast off Angola. The northern part of the Angolan shelf is characterised by large areas of fine to coarse sand. Outside the Congo River in the north of Luanda, silt appears as the main component of the soil. These areas are interrupted by beds of rocks, stones and corals. The central part of the Angolan shelf is characterised by alternating fields of mud and fine coarse sand, whereupon silt and clay are dominating. Rocky bottoms are mainly found in the centre of the shelf. Travelling southwards to the Cunene River estuary the bottom is level, consisting of clay, silt and also fine to coarse sand in the region north of 15°S. In the south, the bottom deeper than 100 to 200 m is rough and untrawlable (Bianchi 1992)

# 1 Introduction

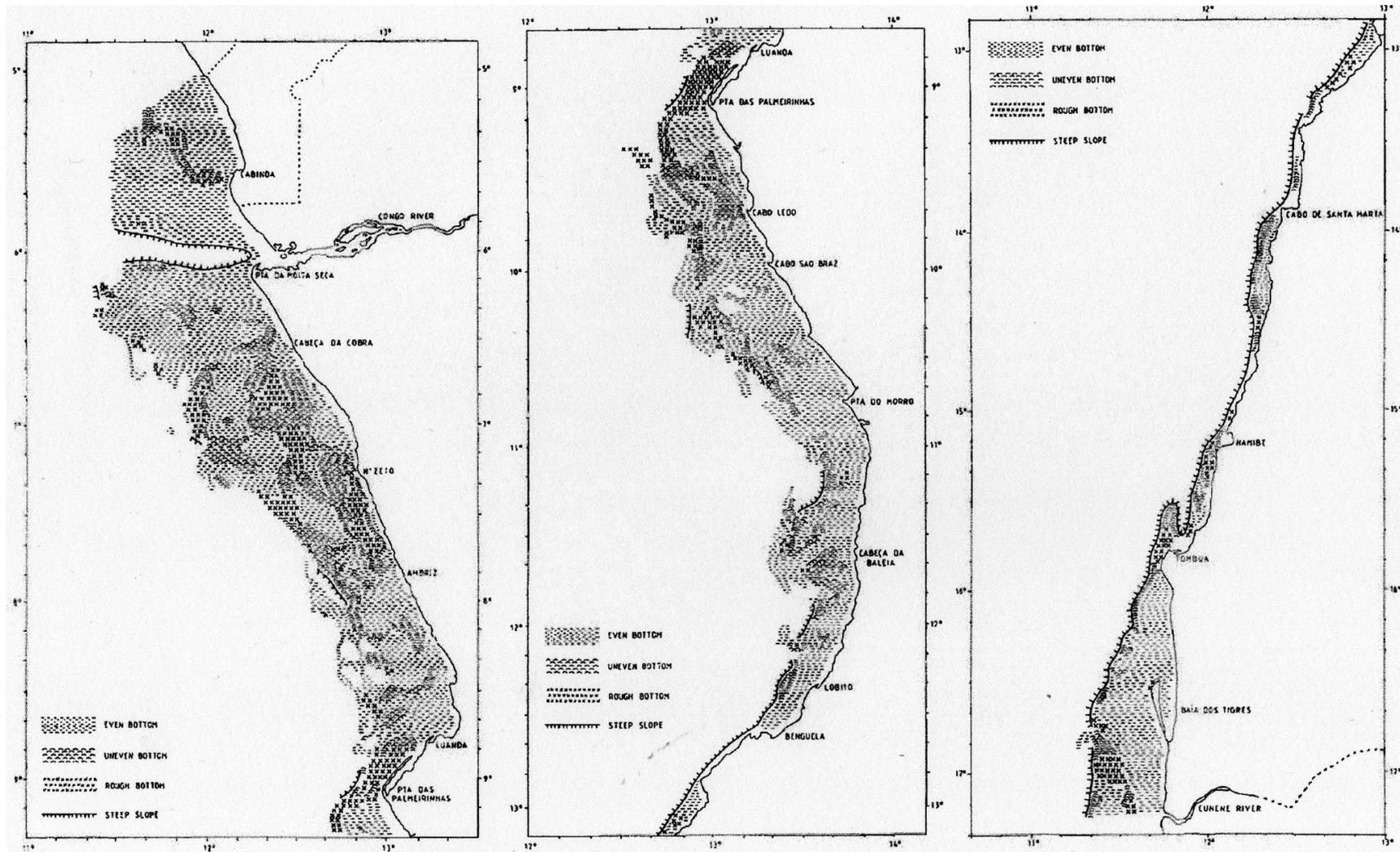
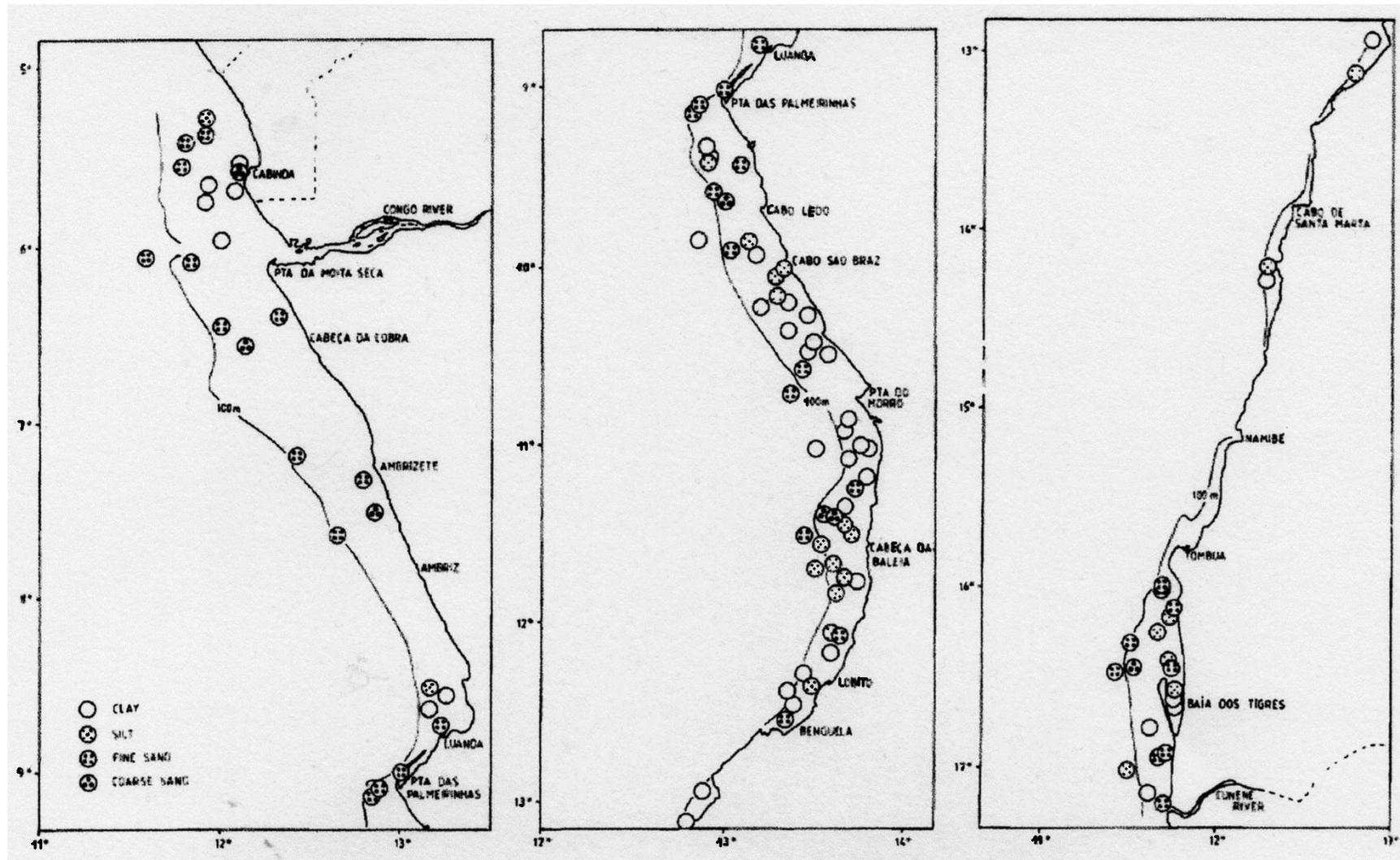


Fig. 2: Shelf bottom structure inferred from echograms along the Angolan coast (Bianchi 1992, redrawn from Strømme & Sætersdal 1991).

# 1 Introduction

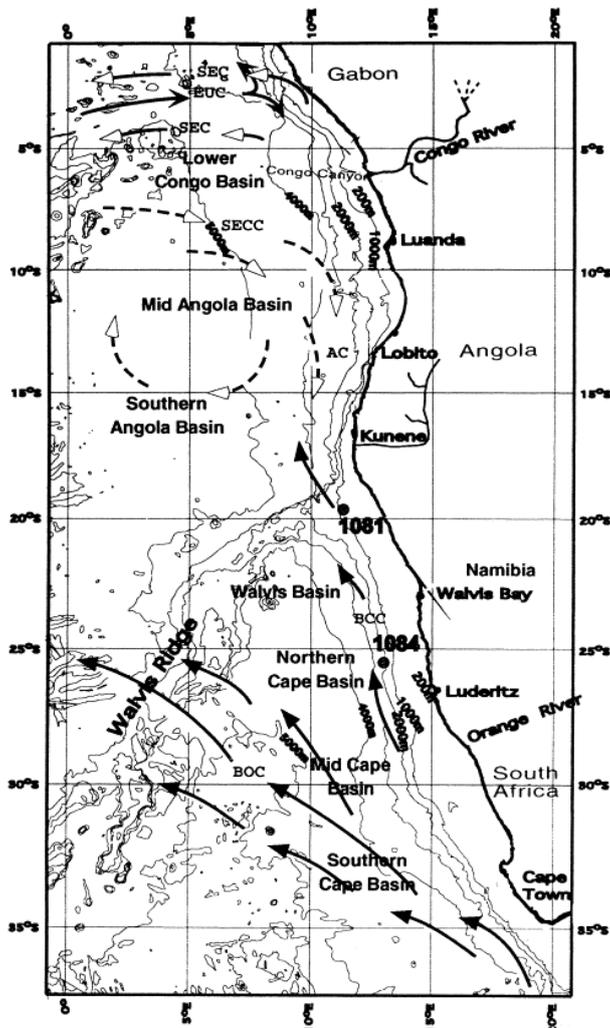


**Fig. 3:** Stations surveyed by Bianchi (1992) in February and March 1989, indicating the types of sediment along the Angolan coast; empty circle: clay, circle with 3 dots: coarse sand, circle with 4 dots: fine sand, circle with 5 dots: silt.

### 1.2.3 Current dynamics

The Angolan coast is mainly influenced by the Angola Current representing the eastern part of a cyclonic gyre that is centred around 1°S and 4°E and is driven by the South Equatorial Countercurrent in the Atlantic Ocean (Figure 4).

At about 16°S, the southward-flowing Angola Current converges with the northward-flowing Benguela Current forming the Angola-Benguela Front (ABF) (Hogan 2010) that sharply separates the nutrient-poor warm water of the Angola Current from the nutrient-rich cold water of the Benguela Current and thus, represents a transition zone between the more typical ecosystem in the north and the upwelling-driven ecosystem in the south (Lass *et al.* 2000).



**Fig.4:** Marine currents in the Southeast Atlantic Ocean at the West African coast; AC: Angola Current, BCC: Benguela Coastal Current, BOC: Benguela Oceanic Current, EUC: Equatorial Undercurrent, SECC: South Equatorial Countercurrent, SEC: South Equatorial Current (Lin & Chen 2002).

The coastal poleward-flowing Angola Current extends from below the surface to 200 m depth in which current speeds of 70 cm/s at the surface and 88 cm/s subsurface were measured during late summer. This poleward undercurrent is distinctive of near bottom water over the shelf and extends westwards of the shelf-break. The subtidal currents over the shelf are dominated by coastal trapped waves. A net poleward flow of 5-8 cm/s or about 5 km/d was noticed. The Angola Current occasionally reaches the surface, resulting in episodes of poleward flow at the surface (Steele *et al.* 2009).

Between the latitudes of 5° and 15°S generally low values of the windstress curl were observed by Lass *et al.* (2000).

### **1.3 Objective**

The aim of this study is the investigation of the composition and distribution of the macrozoobenthic communities along a latitudinal gradient on the shelf off Angola. In the subsequent paragraph, the procedure including materials will be described.

## 2 Material and methods

### 2.1 Material

Benthic samples for this study were taken along a transect at the Angolan coast between Namibia and Luanda (Figure 5) on a cruise of the German research vessel Maria S. Merian (MSM 18/4) from July 29<sup>th</sup> to August 03<sup>rd</sup> 2011.

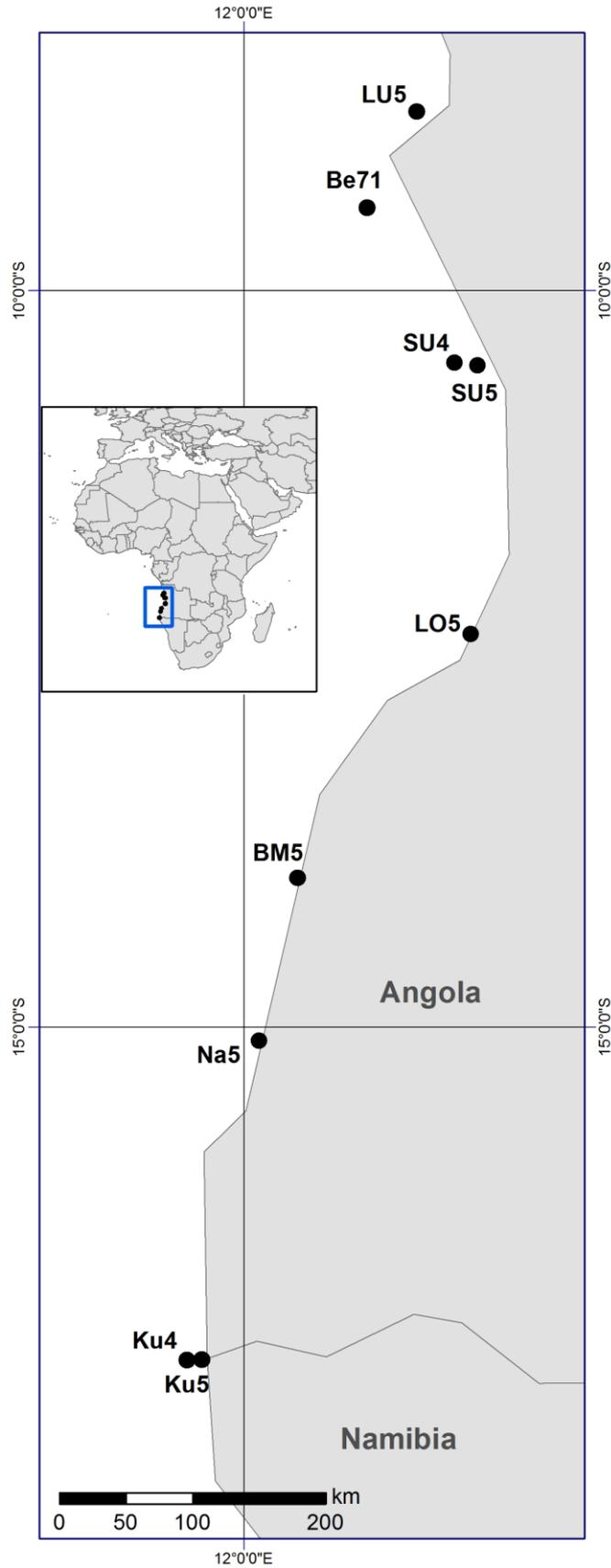
### Sampling

Triplicate samples (haul 1, 2 and 3) were taken with a 0.1 m<sup>2</sup> van Veen grab at each of 9 stations from 28 m to 102 m depth. Additional dredge hauls were taken for collection of mobile, larger or rare species. All samples were sieved through a 1 mm<sup>2</sup> screen and animals were preserved on board in 4% buffered formaldehyde.

A list of all analysed samples including the number of hauls and dredge hauls is given in Table 1. The biotic and abiotic features of the sediment are provided in Table 2.

MSM ID	Station name	Latitude	Longitude	Date of sampling	Depth [m]	H1	H2	H3	D	Total
825	LU5	08°46,980'S	13°10,020'E	03.08.2011	80	1	1	1	3	5
819	Be71	09°26,070'S	12°49,870'E	02.08.2011	102	1	1	1	2	6
813	SU5	10°30,500'S	13°34,600'E	01.08.2011	28	1	-	-	-	1
814	SU4	10°29,380'S	13°25,350'E	01.08.2011	60	1	-	-	1	2
812	LO5	12°20,000'S	13°32,000'E	01.08.2011	60	1	1	1	2	5
807	BM5	13°59,477'S	12°21,690'E	31.07.2011	48	1	1	1	1	4
802	Na5	15°05,750'S	12°06,300'E	30.07.2011	62	4	-	-	-	4
851	Ku4	17°15,840'S	11°36,910'E	15.08.2011	102	1	1	1	2	5
852	Ku5	17°15,700'S	11°43,000'E	15.08.2011	39	1	1	1	1	4

**Tab.1:** List of all analysed samples including the total number of samples, the location of each station, date of sampling and depth; MSM: Maria S. Merian, H: haul, D: dredge haul.



**Fig.5:** Physical map showing the locations of all sampling stations along the Angolan coast (created with GIS ArcMap, version 10).

Station	Organic part [%]	GS [ $\mu\text{m}$ ] 50%	O <sub>2</sub> [ml/l]	Salinity	Description of the substrate
LU5	15,04	7	1,08	35,7	grey/brown soft organic silt
Be71	5,51	59	1,3	35,7	hard clay-containing silt, some shell detritus
SU5	2,18	58	2,3	35,7	muddy and fine sand with shell detritus and diatoms
SU4	n.m.	n.m.	n.m.	35,6	lithoidal, coarse sand
LO5	11,96	14	1,04	35,7	brown/grey soft mud
BM5	2,37	87	1,36	35,7	dark grey muddy fine sand
Na5	8,1	14	1,28	35,6	muddy mixed sand with diatoms; first 3 cm oxidized; below: grey/black colour
Ku4	8,83	18	0,66	35,5	soft dark brown/black silt, intense H <sub>2</sub> S smell
Ku5	9,88	23	0,82	35,5	soft black silt, 2 mm brown coating, intense H <sub>2</sub> S smell

**Tab.2:** Values of the abiotic and biotic features of each sampling station and sediment characteristics; GS: grain size, n.m.: not measured.

## 2.2 Methods

### 2.2.1 Sample processing

In laboratory, 36 samples, fixed in 4 % formalaldehyde sea water, were washed over a 0,5 mm mesh and conducted with a stereomicroscope with 10-40x magnification. The organisms were determined with different literature, also using the internet (e.g. <http://marinespecies.org/>). All specimens were identified to the lowest taxonomic level whenever possible. For preservation, the organisms were fixed in 95% ethanol and glycerol. The biomass was determined by a special accuracy weighing machine (Analytical balance Cubis<sup>®</sup> MSA225S-000-DA, Sartorius GmbH). Furthermore, the key species were photographed (AxioVision: version 4.8.2.0).

### 2.2.2 Statistical analysis

Multivariate community analysis was done using Primer (version 6) with the whole abundance. To detect possible differences in assemblage composition between habitats, this analysis was carried out on square-root transformed and abundance data. Bray-Curtis index and group average linkage were performed for cluster analysis and non-metric multidimensional scaling (MDS) ordination.

### 3 Results

This paragraph is divided in different categories including diversity, community analysis, biomass and abundance as well as key species.

The stations in all diagrams are sorted in the order of the south-north-gradient.

It has to be taken in account that the effort of sampling at the stations was different, so that only one haul was respectively analysed from Na5, SU4 and SU5, while from station SU4 a dredge haul was also present. That is why the standard deviations are missing in the diagrams. From the other stations (Ku4, Ku5, BM5, LO5, Be71, LU5) 3 hauls and at least one dredge haul were analysed.

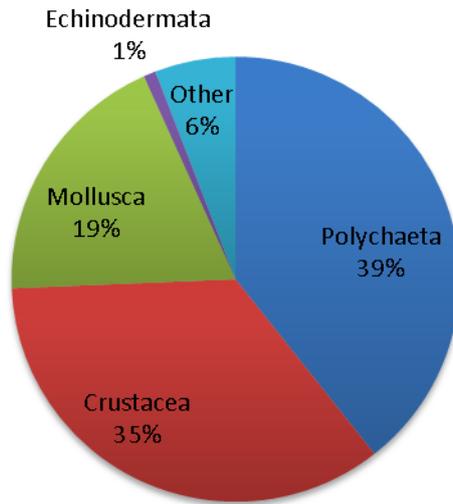
#### 3.1 Diversity

A total of 343 taxa were encountered that were sorted in the main groups Polychaeta, Crustacea, Echinodermata, Mollusca and Other (see appendix). The number of taxa within the main groups is provided in Table 3.

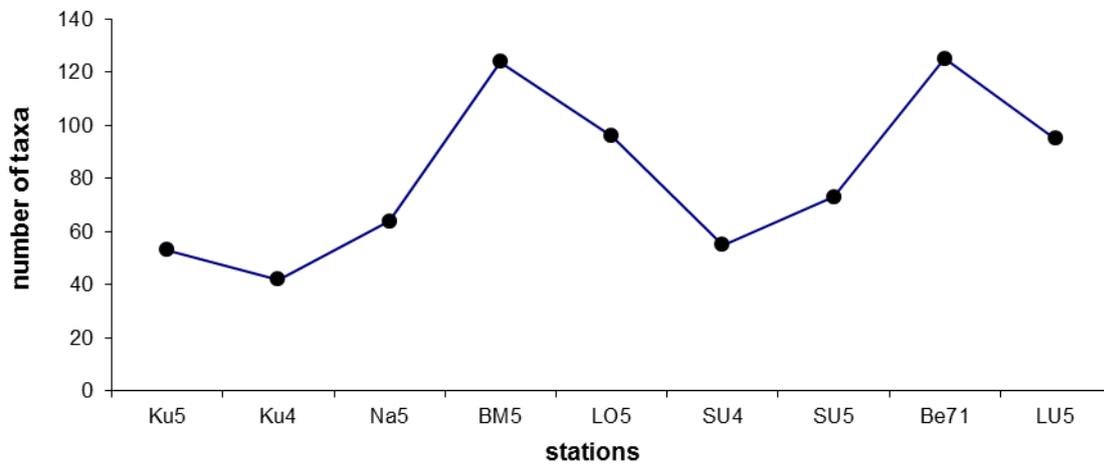
Main group	Taxa
Polychaeta	135
Crustacea	120
Mollusca	65
Echinodermata	3
Other	20

**Tab.3:** Overview of the number of taxa within each main group.

The macrobenthic population on the shelf off Angola is dominated by the polychaetes. They contributed 39%, followed by crustaceans (35%), molluscs (19%) and Other (6%). The minority is represented by the echinoderms with 1% (Figure 6). Figure 7 graphically illustrates the variation in the number of taxa along the stations.

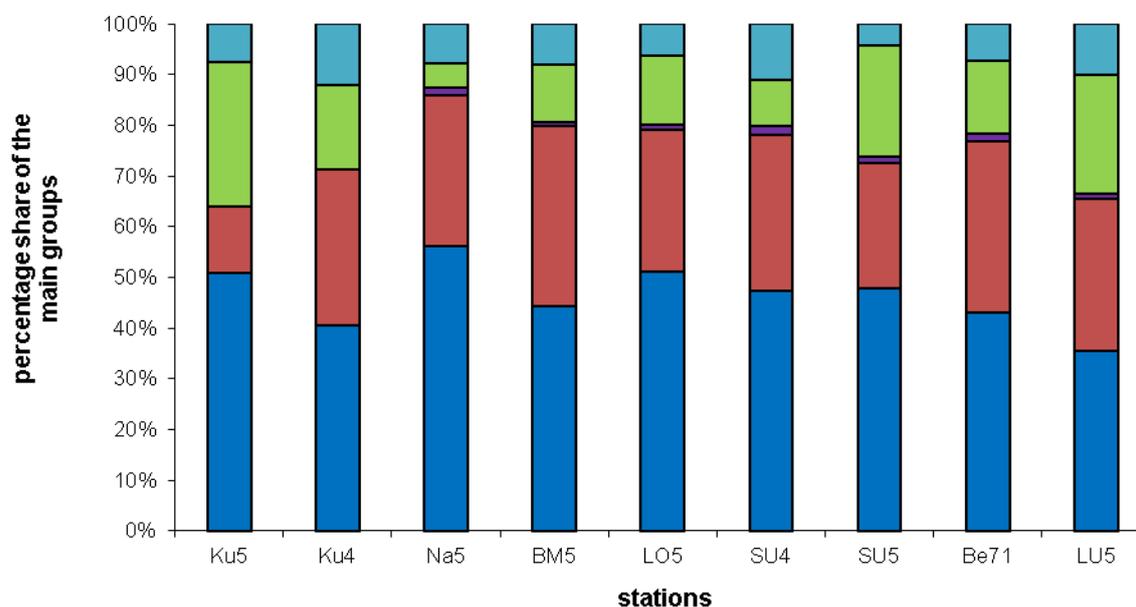


**Fig.6:** Composition [%] of the investigated area.



**Fig.7:** The total number of taxa at each sampling station.

The percentage share of the main groups is illustrated graphically in Figure 8. The exact values are provided in Table 4, including the total number of taxa at each station as well as the number of taxa within each main group.

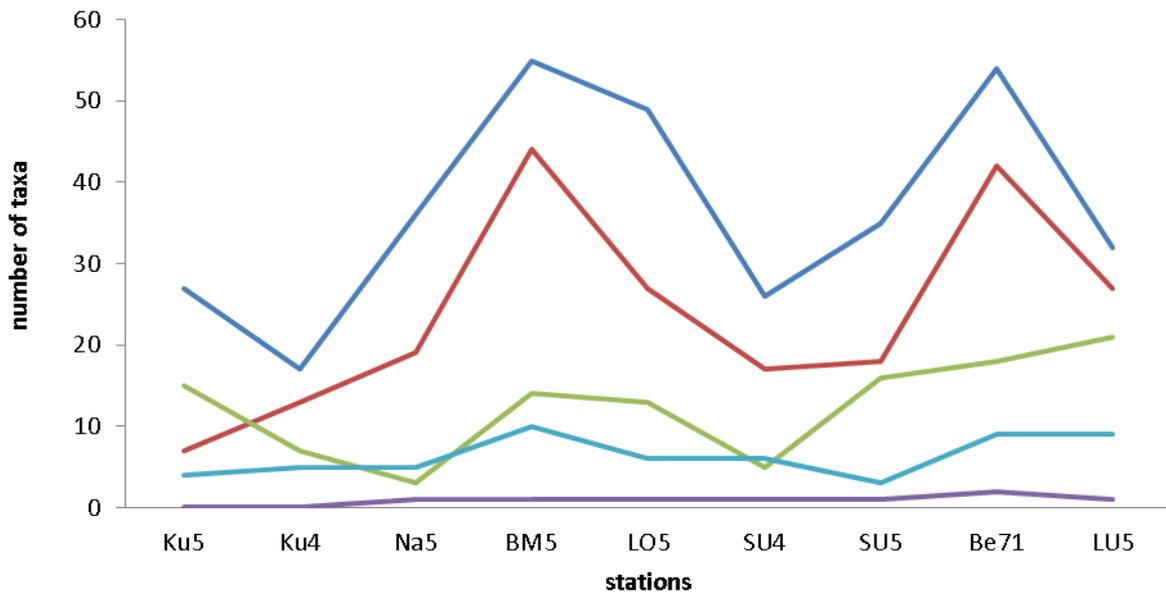


**Fig.8:** Percentage share of the main groups at each station; blue: Polychaeta, red: Crustacea, purple: Echinodermata, green: Mollusca, turquoise: Other.

	Ku5	Ku4	Na5	BM5	LO5	SU4	SU5	Be71	LU5
<b>Polychaeta</b>	27	17	36	55	49	27	35	54	32
<b>Crustacea</b>	7	13	19	44	27	16	18	42	27
<b>Echinodermata</b>	0	0	1	1	1	1	1	2	1
<b>Mollusca</b>	15	7	3	14	13	5	16	18	21
<b>Other</b>	4	5	5	10	6	6	3	9	9
<b>Total number</b>	53	42	64	124	96	55	73	125	90

**Tab.4:** Number of taxa within the main groups of each sampling station as well as the total number.

The values of each main group vary significantly along the stations (Table 4). An alternating trend is remarkable (Figure 7). The most abundant group in all stations is represented by the polychaetes (17 to 54 taxa), whereupon the echinoderms exhibit the lowest values of 0 to 2 taxa. The trend of each main group can be seen in Figure 9.



**Fig.9:** Diversity trends of all groups; blue: Polychaeta, red: Crustacea, purple: Echinodermata, green: Mollusca, turquoise: Other.

It can be noticed that all curves fluctuate, except that one of the echinoderms. The curve shapes of Polychaeta, Crustacea and Mollusca appear to be similar, whereat the diversities of the polychaetes and mollusks reveal the greatest fluctuations.

### **Biodiversity indices**

There is a variety of indices for measuring biodiversity that have been described by Kahn (2006). In this study, the Shannon- and the Margalef index were used. They are shortly summarised in the next paragraphs followed by the results for each station (Figure 5). The results of the dredge samples were not incorporated in the calculation.

#### a) Shannon index

The Shannon index reflects the complexity of biological interactions in a community and is the very widely used index for comparing diversity between various habitats (Clarke & Warwick 2001). The so-called Shannon-Weaver equation ( $H'$ ) describing this model was derived from Shannon's information theory (Oliver *et al.* 2011).

For each station, the Shannon index was calculated using the following formula:

$$H' = - \sum p_i \cdot \log_2$$

with  $p_i = n_i / N$ .

N indicates the total number of species,  $n_i$  the species and  $p_i$  the proportion of individuals belonging to the  $i$ -th species in the dataset of interest - the higher the value, the greater the diversity.

b) Margalef index

This index is also a measure of species diversity that has a very good discriminating ability and is sensitive to sample size (Kahn 2006). Furthermore, it can be interpreted as the ratio between the maximum number of interspecific interactions and the maximum number of intraspecific interactions (Giavelli *et al.* 1986). It is calculated from the total number of present species and abundance or total number of individuals:

$$d = (s-1)/\log N,$$

where  $s$  describes the number of species and  $N$  indicates the total number of individuals in the sample (Margalef 1958).

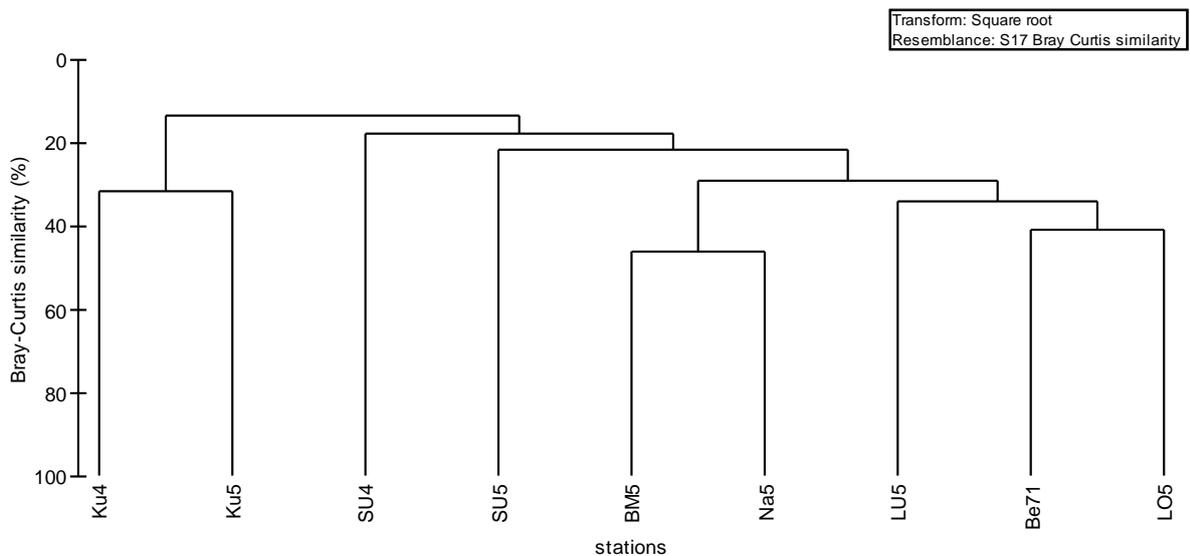
Station	Shannon index	Margalef index
LU5	4,10	28,83
Be71	5,44	36,86
SU5	4,23	18,53
SU4	3,96	16,60
LO5	4,93	29,50
BM5	2,35	31,43
Na5	3,38	13,74
Ku4	3,07	13,33
Ku5	2,75	13,14

**Tab.5:** The calculated values of the Shannon index and Margalef index for each station.

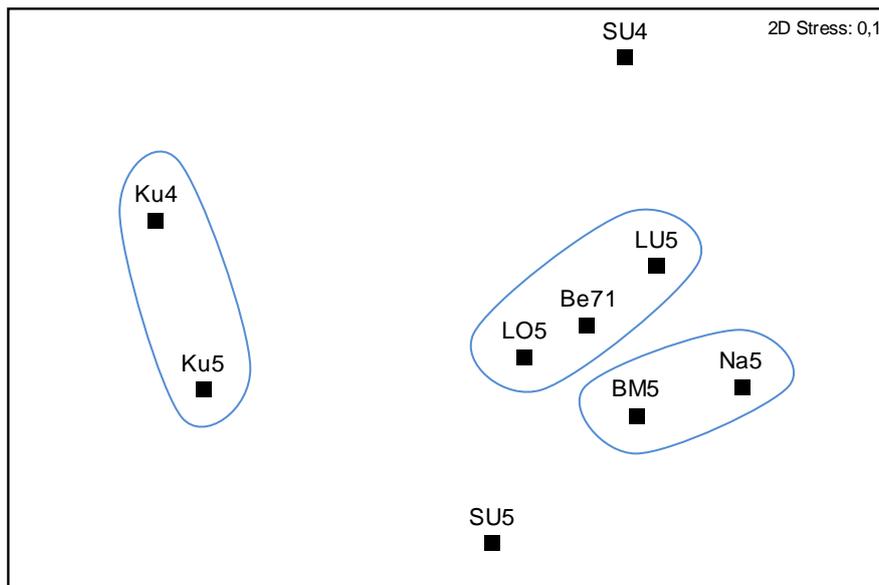
The calculated values of the Shannon index vary between 2,31 (station BM5) and 5,44 (station Be71). The values of the Margalef index range from 13,33 (station Ku4) to 36,86 (station Be71).

### 3.2 Community analysis

Multilinkage cluster analysis by group average linkage was applied for grouping the species into various clusters at different similarity levels adopting Bray Curtis similarity index. The results are shown graphically as a dendrogram and in a MDS-plot. All determined organisms, from both, hauls and dredge samples, the quantity and the stations were incorporated in the calculation.



**Fig.10:** Hierarchical, agglomerative clustering of square root-transformed macrobenthos data of the abundance of all stations using group-average linking on Bray-Curtis similarities (%).



**Fig.11:** Multidimensional scaling ordination for square root-transformed macrobenthos data of the abundance based on Bray-Curtis similarities (stress=0.1).

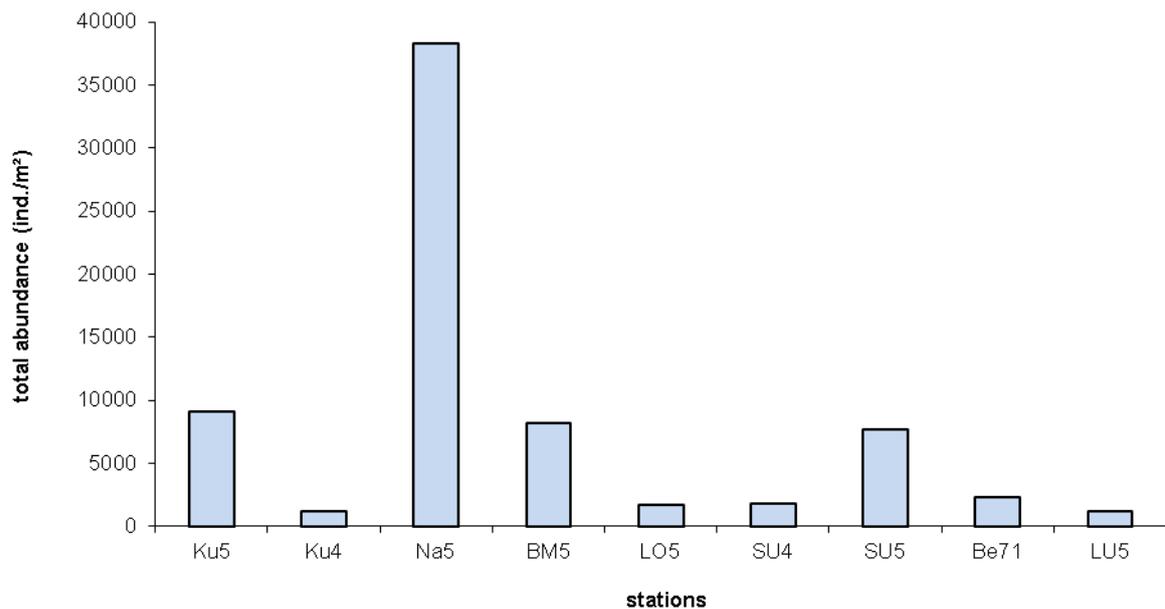
Clusters (Figure 10) occur over a relatively small range of similarities ( $\sim 10 - 35\%$ ). At about 10% the sites are divided into two clusters, and at 20% or rather 30% into two main groups. One group comprises a cluster of the two stations in the south (Ku4 and Ku5), the other group implies the stations located to the north of the first group containing the clusters of the single stations SU4 and SU5 as well as a site of the stations LU5, Be71 and LO5.

The sites in stations BM5 and Na5 show highest similarity to each other ( $\sim 45\%$ ), whereas the site in station SU4 is most dissimilar. The MDS ordination (Figure 11) illustrates the same result as a 2D-diagram.

### 3.3 Abundance and Biomass

The abundance and biomass are both shown graphically (Figures 12, 14) containing the total values of each sampling station. The exact values of each group are provided in Table 6 (abundance) and Table 7 (biomass). The variations of each group along the coast are additionally illustrated graphically in Figure 13 (abundance) and Figure 15 (biomass).

#### Abundance

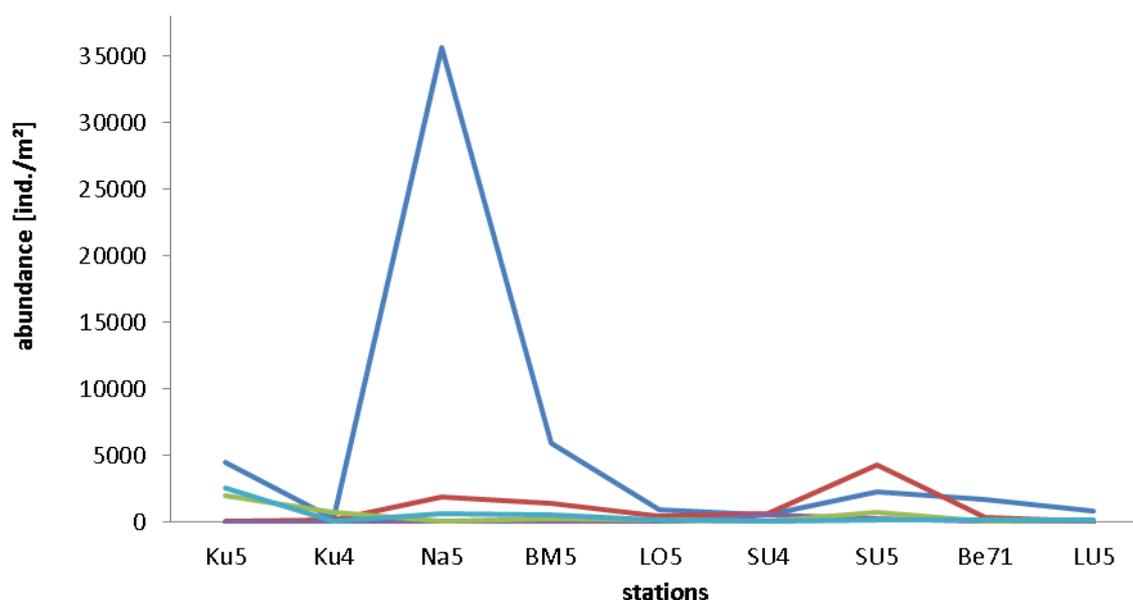


**Fig.12:** Total abundance [individuals/m<sup>2</sup>] of each sampling station.

### 3 Results

	Ku5	Ku4	Na5	BM5	LO5	SU4	SU5	Be71	LU5
<b>Polychaeta</b>	4.501	277	35.662	5.885	931	499	2.258	1.678	858
<b>Crustacea</b>	50	122	1.922	1.443	442	668	4.328	307	99
<b>Echinodermata</b>	0	0	21	95	4	525	221	81	4
<b>Mollusca</b>	1.939	766	42	218	114	24	735	69	111
<b>Other</b>	2.605	23	685	551	173	77	148	179	151
<b>Total</b>	9.095	1.188	38.332	8.192	1.664	1.793	7.690	2.314	1.223

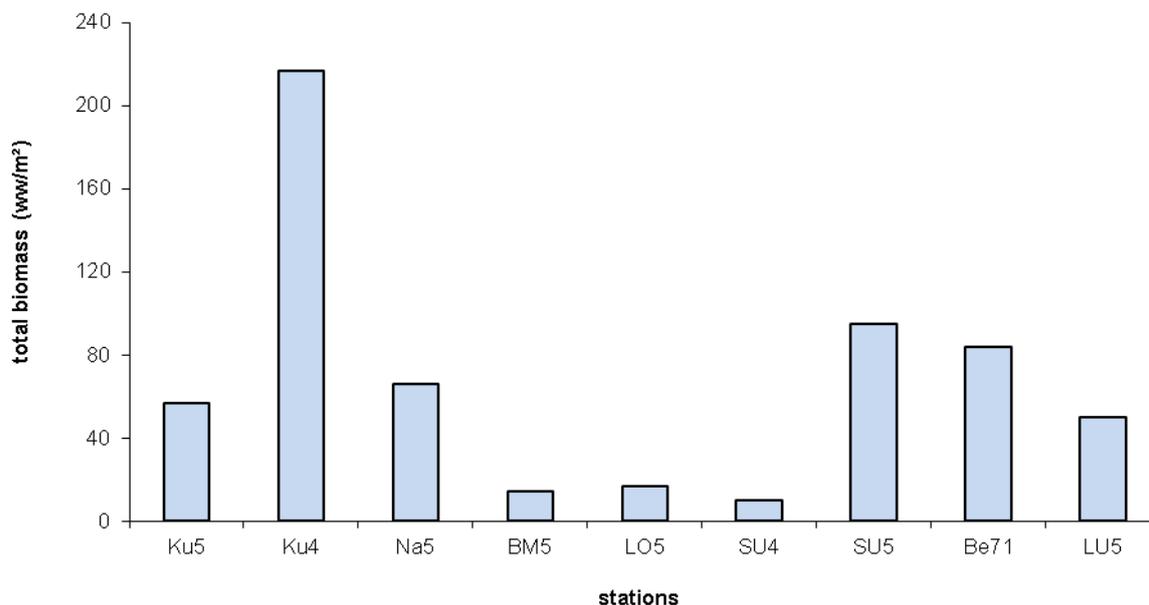
**Tab.6:** Abundance [individuals/m<sup>2</sup>] of each main group at each station, including single and total values.



**Fig. 13:** Abundance of each main group; blue: Polychaeta, red: Crustacea, purple: Echinodermata, green: Mollusca, turquoise: Other.

The values of the total abundance vary between 1.188 ind./m<sup>2</sup> (Ku4) and 38.332 ind./m<sup>2</sup> (Na5). The polychaetes reveal the highest abundance of 35.662 ind./m<sup>2</sup> at station Na5. This is also illustrated with the highest peak of all curves in Figure 13, whereupon the other curves are considerably flatter. The lowest abundances of 4 ind./m<sup>2</sup> were determined for the echinoderms at stations LO5 and LU5. At the southern stations Ku4 and Ku5, they were not found.

## **Biomass**

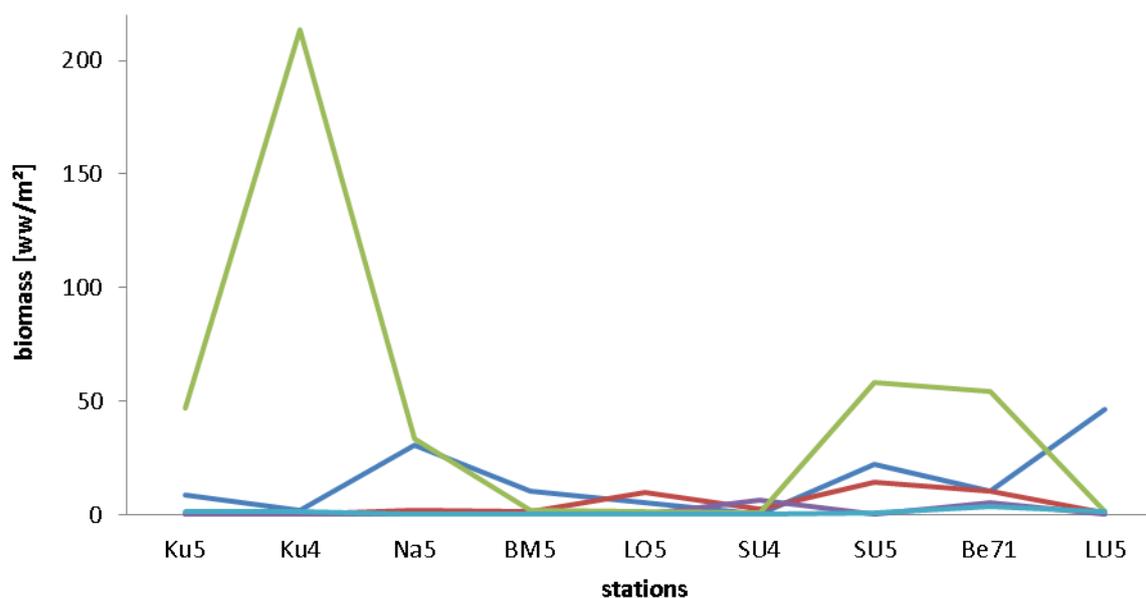


**Fig.14:** The total averaged biomass [wet weight in g/m<sup>2</sup>] at each sampling station.

	Ku5	Ku4	Na5	BM5	LO5	SU4	SU5	Be71	LU5
<b>Polychaeta</b>	8,59	1,68	30,71	10,30	5,07	0,40	22,26	10,21	46,49
<b>Crustacea</b>	0,05	0,07	1,71	1,66	9,73	2,45	14,09	10,16	0,90
<b>Echinodermata</b>	0	0	0,001	0,04	0,29	6,55	0,003	5,13	0,08
<b>Mollusca</b>	46,72	213,54	33,41	1,99	1,54	0,57	58,25	54,30	1,34
<b>Other</b>	1,34	1,55	0,19	0,45	0,18	0,1	0,60	3,82	1,41
<b>Total</b>	56,71	216,85	66,01	14,43	16,81	10,06	95,20	83,63	50,22

**Tab.7:** The averaged biomass [wet weight in g/m<sup>2</sup>] of each main group at each station, including single and total values.

There is a quite similar trend for biomass (Figure 14). The highest value of about 216,85 g can be found at station Ku4. The lowest value of 10,06 g is provided at station SU4 (Table 7). In the south, the biomass was particularly influenced by the mollusks that were very frequent at station Ku5 (Figure 15, Table 6) and indicates the highest biomass value of 213,54 g/m<sup>2</sup>. In the same region, the echinoderms were not present. They also contributed the lowest biomass of 0,001 g/m<sup>2</sup> (station Na5). Furthermore, the values of the crustaceans are very low in the same area compared to the values of the northern stations, except for LU5.



**Fig.15:** Average biomass of all main groups; blue: Polychaeta, red: Crustacea, purple: Echinodermata, green: Mollusca, turquoise: Other.

### 3.4 Profiles of the key species

Key or keystone species can be defined as species that are dominant in ecosystems and which have large effects on them and their communities (Bengtsson 1997). On the following pages, the key species (Table 8) are presented in a summarised profile, describing their distribution, morphology, nutrition, reproduction and habitat preferences.

The profiles are arranged according to their taxonomic order and within alphabetical order.

Key species	Class	Station	Abundance [ind./m <sup>2</sup> ]
<i>Nuculana bicuspidata</i>	Bivalvia	Ku5/Ku4	382/217
<i>Nassarius vinctus</i>	Gastropoda	Ku5/Ku4	798/529
<i>Cossura coasta</i>	Polychaeta	Ku5	3389
<i>Diopatra neapolitana</i>	Polychaeta	LU5	294
<i>Owenia sp.</i>	Polychaeta	BM5/Na5	2682/9790
<i>Prionospio sp.</i>	Polychaeta	Na5	10200

**Tab.8:** List of all key species arranged according to their class including their locations and their abundance [individuals/m<sup>2</sup>].

### 3.4.1 Bivalvia

#### ***Nuculana bicuspidata* (Gould, 1845)**



**Fig.16:** *Nuculana bicuspidata* from station Ku4.

- Distribution:** This bivalve lives from Mauritania to Angola, but was also found near Velddrif (Namibia) and Cape Cross (South Africa) (Kensley 1985).
- Morphology:** The shiny white shell is elongated, equivalve, asymmetrical, thick-walled, short but rostral elongated posteriorly and rounded anteriorly. The posterior surface is strongly ribbed. The ribs are concentric at the front end, where they are interrupted by a strong incision. The width of the shell ranges from 3 mm to 2 cm (Mangalo 2004).
- Nutrition:** *N. bicuspidata* is a typical suspension feeder and filters detritus (Michel *et al.* 2011).
- Reproduction:** This information is not published.
- Habitat:** The shell is a typical sand-dweller (Kensley 1985) that lives close to the sediment-water interface in organic-rich, fine-grained material (Michel *et al.* 2011).

### 3.4.2 Gastropoda

#### *Nassarius vinctus* (Marrat, 1877)



Fig.17: *Nassarius vinctus* from station Ku4.

- Distribution: This gastropod occurs from East London to Namibia, particularly on continental shelves in 35 to 150 m depth (Kilburn & Rippey 1982).
- Morphology: The dark cream-coloured or whitish shell is ovate, rather thick-walled and relatively wound with deep sutures and an inner lip forming a very thin callus. A thin outer lip and a wide siphonal are also present. The body size ranges from 1 mm to 3 cm.
- Nutrition: *N. vinctus* is an active scavenger (Herbert & Comptom 2007).
- Reproduction: There is an annual episodic breeding season from March to August. The fertilisation of the genus *Nassarius* is external. About 6.000 egg capsules, each one containing about 200 eggs, are attached to stones and rocks. The larvae develop in the water column (Marine Ecological Surveys Limited 2008).
- Habitat: This species typically lives on the sediment surface of soft bottoms (Kilburn & Rippey 1982), especially on muddy gravels (Herbert & Comptom 2007), but also on rocks and boulders (Marine Ecological Surveys Limited 2008).

### 3.4.3 Polychaeta

#### *Cossura coasta* (Kitamori, 1960)



**Fig.18:** *Cossura coasta* (Kitamori 1960) without the posterior part from station Ku5.

- Distribution:** *C. coasta* occurs worldwide from 1 m to 2400 m depth (Rouse & Pleijel 2001).
- Morphology:** The body consisting of a distinct muscular thoracic region and a more fragile `abdomen´ is small (less than 10 mm) and can possess about 100 segments. Living animals are translucent with pale tan or brown tinting (Rouse & Pleijel 2001). The dorsal branchia is a typical characteristic arising from setiger 3 (Day 1963).
- Nutrition:** *Cossura* is a deposit-feeder and uses its unique nuccal tentacles by opening the buccal cavity widely and placing them on the sediment surface (Rouse & Pleijel 2001).
- Reproduction:** This polychaete appears to be gonochoric. Little is known about the reproduction, e.g. whether they copulate or brood larvae in any way (Rouse & Pleijel 2001).
- Habitat:** *C. coasta* is non-tubicolous and burrows in shallow marine sediments, but is more common in mixed sand and mud sediments in the deep sea (Rouse & Pleijel 2001).

**Diopatra neapolitana Delle Chiaje, 1841**

**Fig.19:** *Diopatra neapolitana* Delle Chiaje, 1841 from station LU5.

A: anterior body with head, dorsal-lateral view

B: dorsally head with antennae and palps

- Distribution:** *D. neapolitana* is a cosmopolitan species that has been recorded in the Mediterranean, the Red Sea, the Eastern Atlantic Ocean and Indian Ocean (Rouse & Pleijel 2001).
- Morphology:** The body size varies from a few mm to several metres, but around a few cm is common. The majority is tubicolous. Living animals are iridescent and show characteristic red or brown pigmentation patterns, often as transverse segmental bands. The oval prostomium bears a pair of rounded or elongated frontal lips anteriorly as well as a pair of antero-lateral palps and 3 more posterior antennae. Sexual dimorphism is present (Rouse & Pleijel 2001).
- Nutrition:** This polychaete feeds as carnivore, herbivore and scavenger (Rouse & Pleijel 2001).
- Reproduction:** *D. neapolitana* is gonochoric and releases its gametes in the water column where the larvae develop (Rouse & Pleijel 2001).
- Habitat:** This species inhabits intertidal mudflats and shallow subtidal transitional waters and is capable of constructing tubes produced by a secreted layer that is adhered by sand particles, fragments of solid parts from other animals, e.g. shells, and algae (Pires *et al.* 2012).

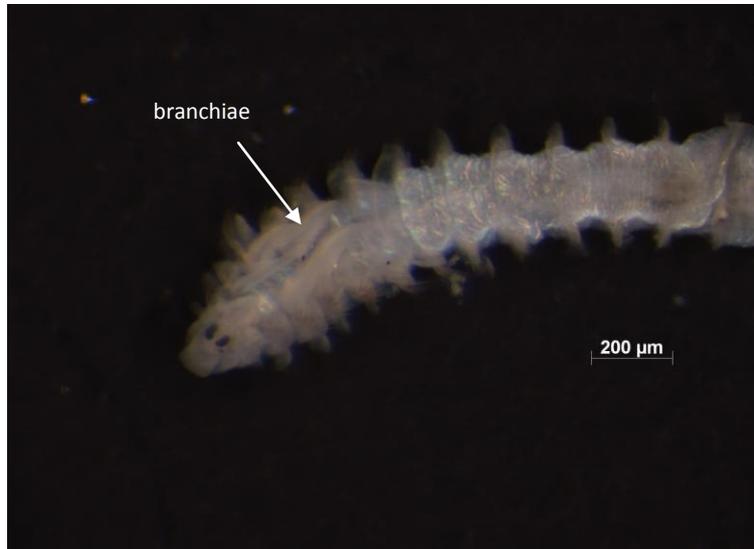
**Owenia sp.**



**Fig.20:** The anterior part of *Owenia* sp. with head from station BM5.

- Distribution:** *Owenia* sp. occurs worldwide to around 200 m depth, but also lives in some abyssal regions (Rouse & Pleijel 2001).
- Morphology:** The body is elongate and cylindrical with between 20 and 30 segments. The length ranges from less than 10 mm to more than 100 mm. The head comprises the prostomium and peristomium, apparently with no associated segments. The prostomium bears a multilobed 'crown' (Rouse & Pleijel 2001).
- Nutrition:** This polychaete is a surface deposit and suspension feeder (De Santa-Isabel *et al.* 1998).
- Reproduction:** *Owenia* is gonochoric. The females shed an average of 70.000 eggs in the water column (external fertilisation) from May to June each year. The larvae develop in the pelagic (Rouse & Pleijel 2001).
- Habitat:** The polychaete lives in intertidal sediments, constructing distinctive tightly fitting sedimentary tubes that often include sand particles, shell fragments and Foraminifera tests and from which *Owenia* is extremely difficult to extract (Rouse & Pleijel 2001).

**Prionospio sp.**



**Fig.21:** The anterior part of *Prionospio* sp. with head from station Na5.

- Distribution:** This polychaete is distributed worldwide from intertidal to abyssal depths (Rouse & Pleijel 2001).
- Morphology:** *Prionospio* ranges in length from a few mm to several cm, shows a variety of colours and pigmentation patterns, and can easily be recognized by the pair of elongate grooved palps extending from the dorsally head, but is not always present (Figure 21). The branchiae can be pinnate and are limited to the anterior region terminating by about segment 15. The prostomium is usually a narrow, ellipsoidal lobe resting on the top of the peristomium. The tip is often rounded (Rouse & Pleijel 2001).
- Nutrition:** The genus is a typically deposit feeder (Rouse & Pleijel 2001).
- Reproduction:** This gonochoric polychaete spawns its gametes in the water column where are brooded - particularly in spring and summer (Rouse & Pleijel 2001).
- Habitat:** *Prionospio* lives in burrows or tubes in sandy and muddy sediments (Rouse & Pleijel 2001).

## 4 Discussion

This paragraph is divided in a) latitudinal gradient describing the results of this study comparing with prior studies at shelves from different areas with regard to diversity, biomass and abundance, b) key species and c) challenge indicating the difficulties determining the organisms.

### 4.1 Latitudinal gradient

In this study, Figure 7 generally shows an alternating trend for the Angolan shelf. There are two peaks, at station BM5 and Be71, whereupon the stations Ku4, Ku5 and SU4 exhibit the lowest diversities. The result of the cluster analysis (Figure 10) and the MDS-Plot (Figure 11) additionally do not show any evidence of a latitudinal gradient, since stations LU5, Be71 and LO5 form a cluster, not including SU4 and SU5.

Some studies also do not reveal a latitudinal cline for marine fauna in shallow waters (Kendall & Aschan 1993, Boucher & Lamshead 1995, Joydas & Damodaran 2008). Hillebrand (2004), who analysed more than 100 studies from around the world, suggested that the gradient is likely weak if detectable at all (Gray 1994).

Kendall & Aschan (1993), e.g., investigated areas of different climates including the coast of Svalbard in the arctic part of Norway, off the temperate coast of the British Islands and the tropical island Java. They figured out that there was no variation in sample species richness. Clarke (1992), Poore & Wilson (1993) and Crame (2000) pointed out that there could be a difference within the hemispheres, since the evidence of latitudinal gradient of decreasing richness from the tropics to Antarctica in the southern hemisphere is less convincing than in the northern hemisphere. Consequently, the latitudinal cline across all taxa in the sea may not to be similar as seen on land (Clarke 1992, Clarke & Crame 1997).

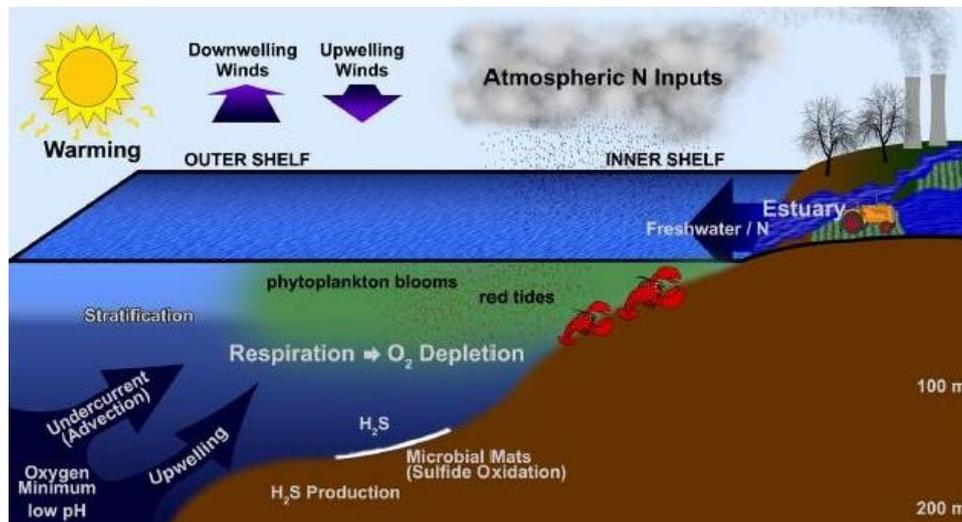
Roy *et al.* (2000) suggested that particularly bivalve species show strong latitudinal diversity gradients. However, Figure 9 reveals also an alternating trend with peaks at stations Ku5, BM5, LO5, SU5, Be71 and LU5. There is only a difference of 6 taxa between the southern station Ku5 and the northern station LU5.

The most southern stations Ku4 and Ku5 in this study are located in the mouth of the

Cunene River representing the northern fringe of an oxygen minimum zone (OMZ;  $O_2 < 0,5$  ml/l) that generally ranges from 80 m to 120 m depth off northern Namibia. In fact, not all organisms are able to cope with hypoxia that influences both the structure and function of benthic communities (Levin *et al.* 2009). Indeed, echinoderms were not found there, but a high abundance of the mollusks *Nuculana bicuspidata* and *Nassarius vinctus*, as well as, the polychaete *Owenia* sp. and specimens of the subclass Oligochaeta could be detected.

‘It is well known that coastal areas with high physical variability such as estuaries contain low diversities’ (Gray *et al.* 1997).

The natural and human-induced hypoxia experienced by benthos in such areas is graphically described in the following illustration.



**Fig.22:** Mechanisms of the formation of hypoxia experienced by zoobenthos along continental shelves (Levin *et al.* 2009).

However, the northern stations do not show this condition. The oxygen content maintains  $>1$  ml/l (Table 2). Thus, a possibly cause for the alternate trend could be the variability in structure and topography of the sediments along the coast.

In fact, Etter & Grassle (1992) found relationships between species and sediment heterogeneity. They suggested that sediment particle size influences the composition of benthic communities and thus, has an important role in determining the number of species within a community. Levin *et al.* (2000) emphasized that depth also has an effect. But surveys, conducted by Gray *et al.* (1997), in both the deepsea and coastal habitats, are

shown to traverse a variety of microhabitats. They hypothesized that sediment heterogeneity may not be an explanation for high species richness in coastal environments. Together with Gray (1994) and Coleman *et al.* (1997), they certainly found high diversity especially in soft sediments in tropical regions. Sanders (1968) also figured out that the number of species in soft-bottoms was higher in the tropics than in the boreal regions.

The soft-sediment benthic macrofauna on the European continental shelf surveyed by Renaud *et al.* (2009) exhibited little evidence of a latitudinal trend that was statistically very small. Nevertheless, a lack of decline in diversity on continental-shelf soft substrates along a latitudinal gradient has also been documented by others (Kendall & Aschan 1993, Ellingsen & Gray 2002).

Contrary to that, Thorson (1952, 1957) showed in different publications that the increases in numbers of species towards the equator is very pronounced in the epifaunas of hard-bottoms, whereupon the number of infaunal species of soft-bottoms did not change in arctic, temperate and tropical seas. These observations agree with the results of Renaud *et al.* (2009).

The results of the survey conducted by Golikov and Scarlato (1973) indicated that, within similar habitats, the distribution of biomass among species is similar at all latitudes. On the shelf off Angola, the biomass varies a lot, both total biomass (Figure 12) and that of each group (Figure 13). The high values in the southern region were primarily caused by the high abundance of the molluscs, particularly of the bivalve *Nuculana bicuspidata*.

So far, many studies have revealed that invertebrate distribution correlates with grain size. However, Snelgrove & Butman (1994) commented that animal-sediment relationships are more variable and that many species are not always associated with a single sediment type. Nichols (1970), Gray (1974) and Etter & Grassle (1992) tried to explain the relationship between sorting and species richness. Well-sorted sediments offer a smaller range of grain sizes and interstitial spaces that may provide fewer niches in contrast to poorly sorted sediments that consequently contain a less diversity.

Comparing the diversity with the grain size in this study, a relationship can be inferred. The measured average grain sizes of 87  $\mu\text{m}$  at station BM5 and 59  $\mu\text{m}$  at station Be71 were the largest in the whole area, whereupon e.g. the oxygen levels were not the highest. Although the bottoms of some other sampling stations were also soft, the grain sizes were

significantly smaller. Thus, these stations showed low diversities compared to the stations BM5 and Be71. Station SU5, with an average grain size of 58  $\mu\text{m}$ , additionally contained a high benthic diversity. However, the low number of species at station SU4 may result from the lithoidal structure of the bottom that is a preferred habitat of epifauna and infauna of rocks, e.g. brittle stars and polychaetes of the family Serpulidae, that were found in this study.

Furthermore, the value of the Shannon index at station Be71 is 5,44 (Table 5). Generally, the values vary between 1,5 and 3,5, and rarely surpasses 4,5. It has been reported that under log normal distribution an amount of  $10^5$  species are required to reach a value  $>5$  (Kahn 2006). All in all, 5 stations exhibit values  $>3,5$ .

At the stations off the big cities Luanda and Lobito, i.e. LU5 and LO5, the diversities of 90 and 96 taxa could be reasoned by anthropogenic influence, e.g. pollution and tourism. Especially, the abundances of 1.223 individuals/ $\text{m}^2$  (LU5) and 1.664 individuals/ $\text{m}^2$  (LO5) are significantly low (Table 6).

An evidence for latitudinal gradient may exist for decapods, especially for brachyuran crabs and penaeids (Natantia). Large diversities were only found at stations from the tropical regions off Angola – 9 taxa of brachyuran crabs at station LU5 (8°S) and 13 taxa of penaeids at station Be71 (9°S). Only a few species were present in the south, except at station Ku4 and Ku5, where low oxygen levels were measured. These decapods usually occur frequently in tropical areas (Bertini *et al.* 2010, Dall 1990, Steele 1988). Bertini *et al.* (2010) figured out that the highest diversity of decapods on the shelf off Brazil was on soft bottoms consisting of very fine sand. The results of this study are agreeable, since the richness of decapods at station SU4, but also SU5, is very low. According to the diversity of brachyuran crabs and penaeids, there may be an evidence for a cline with decreasing distance to the equator, particularly on soft bottoms.

Snelgrove and Butman (1994) also suggested that depth and sediment grain size may act as secondary forces, while it is more likely that the amounts of hydrodynamic energy and the availability of organic matter have more influence on the composition of benthic communities. The energy profile of water above the sediment surface determines the particle size, which in turn has an effect on burrowing organisms. The greater the energy, the higher the velocity, consequently, the more sediment particles will be carried away.

Hence, depth determines the energy of the waves on the bottom. Thus, the effect is the greatest in shallow waters and decreases with increasing distance to the coast (Bergen *et al.* 2001).

In the current study, there is no correlation between depth and diversity. For example, at stations SU4 and LO5 the samples were taken from 60 m depth (Table 1), whereat the number of species ranges from 55 to 96. For stations Ku4 and Be71, both at 102 m depth, there is a difference of 83 species.

The highest abundance of all areas was found at station Na5 (15°S). Many animals such as polychaetes are detritus feeders and use shell detritus and diatoms as food resources. Furthermore, it offers protection. A lot of organisms accumulate there, especially polychaetes like *Prionospio* sp., *Owenia* sp., *Diopatra* sp. and species of the family Cirratulidae.

Most investigations on shelves revealed the predomination of polychaetes (e.g. Joydas & Damodaran 2008) - that is consistent with the result of this study. The hierarchical order of the main groups slightly varies. On the shelf off Angola the main group with the lowest diversity is represented by the echinoderms, just as in other areas (Ellingsen & Gray 2002, Joydas & Damodaran 2008).

For all analyses and comparisons, it must be taken in account that the effort of sampling differed within the stations (Table 1). Consequently, investigating more samples, i.e. more hauls of the stations (e.g. SU4, SU5 or Na5), could significantly change the result of diversity, biomass and abundance.

### 4.2 Key species

The bivalve *Nuculana bicuspidata* (Gould 1945) and the snail *Nassarius vinctus* (Marrat 1877) were most abundant in the south of the sampling area at the mouth of the Cunene River at stations Ku4 and Ku5. This conclusion is consistent with the result of an investigation from 2009 by Zettler *et al.* In addition, a high number of the polychaete *Cossura coasta* was also found at station Ku5. These two mollusks and the polychaete seem to be well adapted to the nearly anoxic and hypoxic conditions in this region, whereupon *C. coasta* also appeared less frequently northwards. This indicates that *C. coasta* may prefer areas with relatively low

oxygen content (<1 ml/l).

The polychaetes *Diopatra neapolitana*, *Owenia* sp. and *Prionospio* sp. showed high frequencies in the northern regions, particularly at stations LU5, BM5 and Na5. All of them burrow in sand and are capable of constructing tubes in which they hide from predators. Therefore, they use sand particles, fragments of other animals, e.g. shells and also Foraminifera tests that adhere to the secreted layer produced by these polychaetes. Especially at station Na5, the sediment contained a lot of diatoms and shell detritus that favour the construction of tubes and also serve as nutritional basis. At station BM5 and LU5, the bottom was very soft consisting of mud with particles of small grain sizes facilitating the process of burrowing. All in all, the appearance with regard to distribution and habitat preferences of these polychaetes agrees with the references of the literature.

### 4.3 Challenge

The most difficult challenge was the identification of the organisms on species level. The majority lacked of segments or characteristic appendices that are required for identification, especially in case of immaturity or physical damages that were e.g. caused while sieving or sampling with the van Veen grab. Another fact is the smallness, since the body lengths of several individuals, predominantly polychaetes, did not exceed a few millimetres.

At last, the identification was rather based on family level than on genus or species level. Consequently, the diversity of each station is expected to be higher than examined so far since the shelf of Angola is an area that is not well-investigated. Thus, the species are not well-known. That makes it more difficult to determine the examined organisms on species level that have been still identified on family level. The time factor played also an important role. Since the identification was often time consuming, the organisms were given so-called working names.

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## 6 Appendix

### 6.1 Materials

In the following all used materials, chemicals and appliances are listed.

Appliances:

Hand sieve:	IOW Ø 180 mm weight: 0,2 kg mesh size: 1 mm
Kautex bottles:	Omnilab 1 litre
Screw neck vials :	clear glass, 40 ml VWR International GmbH
Microscope:	M3Z Discussion Stereomicroscope Stand: Wild Typ 439168 Crossbar: Wild Typ 479887 Lens: Wild M3Z Eyepiece Base: Leica Eyepiece: Wild 10x/21B 445111 ZEISS Microscope Axio Lab.A1 ZEISS SteREO Discovery.V8
Light source:	ZEISS CL 1500 ECO Schott KL 2500 LCD
Camera:	AxioCam ICc3 Resolution: 2028 x 1540 pixels

Scales: Analytical balance Cubis<sup>®</sup> MSA225S-000-DA  
Sartorius GmbH  
Readability: 0.01 mg  
Weighing range: 220 g  
Calibration: International, isoCAL

Forcep set

Photographic trays

Pan of sort

Software: AxioVision: version 4.8.2.0  
Primer: version 6  
GIS, ArcMap: version 10

Chemicals: Formol 35% aqueous solution:  
VWR International GmbH  
902409010

Methylene blue:  
Merck KGaA  
159270

Ethanol 95% (v/v) denaturated TECHNICAL:  
VWR International GmbH  
20827.412

Glycerol 87% technical grade:  
AppliChem GmbH

## 6.2 Data sets

All determined taxa are listed in alphabetical order according to their main group, also indicating their locations. Furthermore, they are arranged according to their order, class, family or phylum.

### 6.2.1 Crustacea

Order	Taxa	Stations
Amphipoda	<i>Ampelisca</i> sp.	Ku4,Na5,BM5,LO5,SU4,SU5,Be71,LU5
	<i>Amphilocheus</i> sp.	SU5
	Amphipoda (red eyes)	Na5,BM5,SU5,Be71,LU5
	Amphipoda B	BM5
	Amphipoda C	BM5
	<i>Apherusa</i> sp.	SU5
	<i>Byopedos</i> sp.	SU5,Be71
	Caprellidae	Na5,BM5,SU4
	cf. <i>Ceradocus</i>	Na5
	<i>Erichthonius</i> sp.	Be71
	<i>Eriopisa</i> sp.	LO5,Be71
	<i>Eriopisa</i> B	LO5
	<i>Eriopisella</i> ?	Ku4
	<i>Eriopisella</i> sp.	LO5
	<i>Eupariambus fallax</i> K.H. Barnard, 1957	Ku4,BM5
	Eusiridae	Be71
	<i>Gammaropsis</i> sp.	Ku5,Na5,BM5,Be71
	<i>Grandidierella</i> sp.	SU4,SU5
	<i>Harpinia</i> sp.	Be71
	<i>Hyperia</i> sp.	SU4
	Isaeidae	Na5,BM5,SU4,Be71
	Isaeidae?	LU5
	<i>Lepidepecreum</i> sp.	BM5
	<i>Leucothoe procera</i> Bate, 1857	Ku5
	<i>Leucothoe</i> sp.	LO5,SU4,Be71
	<i>Leucothoe</i> ?	BM5
	Lysianassidae	Ku4,Na5,BM5
	Lysianassidae?	BM5
	<i>Maera</i> ?	BM5,LO5,SU4
	<i>Melita</i> sp.	SU5
	Melitidae	Ku4
	Melitidae?	LO5
	cf. <i>Metopa</i>	Na5
	Oeditoceridae	Ku4,Na5,BM5,SU4,Be71,LU5
	Oeditoceridae B	Na5
	<i>Orchomene</i> sp.	BM5,SU4
	<i>Periocolodes</i> sp.	Ku4,Na5,BM5
	<i>Photis</i> sp.	BM5,SU5

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<b>Amphipoda</b>	Phoxocephalidae Phoxocephalidae? <i>Phthisica</i> sp. Sthenotoidae? <i>Synchelidium</i> sp. cf. <i>Thaumatoplax</i> <i>Tiron</i> sp.	BM5,LO5,SU4,Be71 BM5 SU5 Na5 SU5 Ku5 SU4
<b>Cumacea</b>	<i>Bodotriia</i> sp. <i>Bodotriia</i> ? Cumacea Cumacea with melanophores <i>Cyclaspis</i> sp. <i>Diastylis</i> sp. <i>Eocuma cadenati</i> Fage, 1950 <i>Eocuma cadenati</i> ? <i>Eocuma calmani</i> Fage, 1928 <i>Eocuma cochlear</i> Le Loeuff & Intes, 1972 <i>Eocuma lanatum</i> Le Loeuff & Intes, 1972 <i>Eocuma</i> sp. <i>Iphinoe</i> sp.	SU5 BM5 Ku5 BM5 BM5 Ku4,Na5,BM5,LO5,SU5,Be71,LU5 BM5 BM5 Be71 SU5 Na5,BM5 Ku5 BM5,SU5
<b>Decapoda</b>	Brachyura A Brachyura A2 Brachyura A3 Brachyura B Brachyura C Brachyura E Brachyura F („Schamkrabbe“) Brachyura K Brachyura L (cf. <i>Parthenope</i> ) Brachyura M Brachyura G (cf. <i>Rochinia</i> ) Brachyura H (Majidae) Brachyura K (Ebalinae) Brachyura N Brachyura O Brachyura P Brachyura Q <i>Callianassa</i> sp. <i>Callianassa</i> juv. <i>Crangon</i> sp. Euphausiacea Euphausiacea juv. cf. <i>Pachygrapsus</i> <i>Galathea</i> sp. <i>Galathea</i> -like <i>Galathea</i> juv. Lobster larva? 2 <i>Macropodia</i> sp. <i>Megamphopus</i> sp. <i>Munidopsis</i> sp. Natantia A	LU5 LO5,LU5 LO5 LO5,Be71,LU5 SU4,Be71 LU5 LU5 LO5 LU5 BM5,Be71,LU5 LU5 BM5,LU5 Be71 LO5 LO5 LO5,Be71 Be71 Ku4,BM5,LO5, SU5,Be71 BM5 Be71 BM5 BM5,LU5 Ku4 BM5,LO5,Be71,LU5 BM5 Na5 BM5 Be71 SU5 SU4 Be71

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<b>Decapoda</b>	Natantia B	LO5,Be71,LU5
	Natantia C	SU4
	Natantia D	LU5
	Natantia E	Be71
	Natantia G	Be71, LU5
	Natantia H	BM5, LO5, Be71, LU5
	Natantia I	LO5, Be71, LU5
	Natantia juv.	LO5
	Natantia K	LU5
	Natantia L	Be71, LU5
	Natantia M	LO5, Be71, LU5
	Natantia N	BM5, LO5, Be71
	Natantia O	BM5, Be71
	Natantia P	LO5
	Natantia Q	Be71
	Natantia S	Be71
	Natantia U	Be71
	<i>Pagurus</i> sp.	Ku5, Ku4, BM5, LO5, SU4, SU5, Be71, LU5
	Stomatopoda	SU5, Be71
<b>Isopoda</b>	Arcturidae	Na5, BM5, SU5
	<i>Cyathura</i> sp.	Na5, BM5, Be71
	<i>Gnathia</i> sp.	LU5
	Isopoda	Ku4, BM5
<b>Mysidacea</b>	Mysidacea	Ku4, SU4, LU5
	Mysidacea juv.	LO5
<b>Sessilia</b>	<i>Balanus</i> sp.	Ku5
	<i>Pollicipes</i> sp.	LU5
<b>Tanaidacea</b>	<i>Apeudes grossimanus</i> Norman & Stebbing, 1886	Be71
	<i>Calozodion</i> ?	SU4
	<i>Hemikalliapseudes</i> sp.	Na5, BM5, LO5
	Tanaidacea	Be71

**Tab. 9:** Determined crustaceans arranged according to the order, and their locations.

### 6.2.2 Echinodermata

Class	Taxa	Stations
<b>Ophiuroidea</b>	<i>Amphiura</i> sp.	Ku5, SU5, Be71
	<i>Ophiura</i> sp.	BM5, LO5, SU4, LU5
-	Echinodermata	Be71

**Tab.10:** Determined echinoderms arranged according to the class, also undetermined echinoderm, and their locations.

## 6.2.3 Mollusca

Class	Taxa	Stations
<b>Bivalvia</b>	<i>Abra</i> sp.	Ku5, Na5, LO5, Be71, LU5
	Bivalvia D	SU5
	Bivalvia: small, white	Ku5
	cf. <i>Lucinoma capensis</i>	Be71, LU5
	cf. Ostreidae	LU5
	<i>Congetica congoensis</i> (Thiele & Jaeckel, 1931)	SU5
	<i>Corbula</i> sp.	LO5
	<i>Costellipitar peliferus</i> (Cosel, 1995)	Ku5
	<i>Cuspidaria</i> sp.	LO5, Be71
	<i>Dosinia</i> sp.	BM5
	<i>Lucinoma capensis</i> (Thiele & Jaeckel, 1931)	Be71
	<i>Macoma</i> sp.	SU5
	Mytilidae	Ku5, SU5, LU5
	<i>Nucula</i> sp.	LU5
	<i>Nuculana bicuspidata</i> (Gould, 1845)	Ku5, Ku4
	<i>Nuculana</i> cf. <i>commutata</i> (Philippi, 1844)	LU5
	<i>Nuculana</i> -like	BM5
	<i>Phaxas</i> sp.	LO5, SU4, LU5
	<i>Pitar</i> sp.	Ku5, BM5, LO5, SU5, Be71
	<i>Solemya</i> cf. <i>togata</i> (Poli, 1791)	BM5
<i>Tellina</i> sp.	BM5	
<i>Thyasira</i> sp.	Ku5	
<b>Cephalopoda</b>	Octopoda	LU5
<b>Gastropoda</b>	<i>Acteon</i> sp.	SU5
	<i>Bufoaria marginata</i> (Gmelin, 1791)	Ku4, Na5
	<i>Bullia skoogi</i> (Odhner, 1923)	Ku5, Ku4
	<i>Cancilla scrobiculata crosnieri</i> Cernohorsky, 1970	Be71
	<i>Clavatula</i> sp.	LU5
	<i>Cylichna</i> sp.	Ku5, BM5, Be71, LU5
	<i>Eulima</i> sp.	Ku5
	<i>Euspira fusca</i> (Blainville, 1825)	LU5
	<i>Euspira grossularia</i> (Marche-Marchad, 1957)	LU5
	<i>Euspira notabilis</i> (Jeffreys, 1885)	LU5
	<i>Fusinus</i> sp.	Na5, BM5
	Gastropoda	Be71
	Gastropoda	Be71
	Gastropoda B	LO5
	Gastropoda B	LU5
	Gastropoda C	SU5
	Gastropoda C (twisted)	Be71
	Gastropoda D	Be71
	<i>Gibberula</i> sp.	BM5
	<i>Jolya letourneauxi</i> Bourguignat, 1877	SU5
	cf. <i>Lippistes cornu</i> (Gmelin, 1791)	LU5
	<i>Nassarius</i> sp.	Ku5, LO5, SU5, Be71
<i>Nassarius vincetus</i> (Marrat, 1877)	Ku5, Ku4	

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<b>Gastropoda</b>	<i>Natica acinonyx</i> Marche-Marchad, 1957	LU5
	<i>Natica bouvieri</i> Jousseau, 1883 (= <i>N. canariensis</i> Odhner, 1931)	BM5
	<i>Natica marchadi</i> Pin, 1992	Ku5
	<i>Natica multipunctata</i> (Blainville, 1825)	LU5
	Nudibranchia	SU5, Be71
	<i>Philine aperta</i> (Linnaeus, 1767)	Ku5, BM5, LU5
	<i>Philine</i> sp.	BM5, LO5, SU4, LU5
	Rissoidae	SU5, LU5
	<i>Strombina descendens</i> (von Martens, 1904)	Be71
	<i>Tectonatica rizzae</i> (Phillipi, 1844)	LU5
	<i>Tectonatica sagraiana</i> (d'Orbigny, 1842)	Ku5, Ku4, LO5, SU5, Be71
	Turridae	Be71
	Turridae?	Ku4, BM5
	<i>Turritella</i> sp.	Ku4, LO5, SU4
Ungulunidae	SU5	
<b>Polyplacophora</b>	Polyplacophora	SU4
<b>Scaphopoda</b>	Scaphopoda	BM5, LO5, Be71, LU5
<b>Solenogastres</b>	Solenogastres	LO5, LU5

**Tab.11:** Determined mollusks arranged according to the class, and their locations.

### 6.2.4 Other

Phylum	Taxa	Stations
<b>Annelida</b>	Oligochaeta	Ku5, Na5, BM5, LO5, LU5
<b>Arthropoda</b>	Pycnogonida	Na5, BM5
<b>Brachiopoda</b>	Brachiopoda	LU5, Be71
	Brachiopoda B	Be71
	<i>Discinisca</i> sp.	BM5
<b>Bryozoa</b>	Bryozoa	Ku4, Na5, BM5, LU5
<b>Chordata</b>	<i>Molgula</i> sp.	Ku4,
<b>Cnidaria</b>	Anthozoa	SU4, Ku4, Na5, BM5, LO5, LU5, Be71
	Anthozoa?	LU5
	Coral	BM5, LU5
	<i>Edwardsia</i> ?	Ku5, BM5
	Hydrozoa	Ku4, Na5, BM5
	<i>Octocorallia</i> soft	Ku4
	Plumose anemone	LU5
<i>Virgularia</i> sp.	SU4, SU5	
<b>Nemertea</b>	<i>Lineus</i> sp.	LO5, Be71, LU5
	Nemertini	Ku5, BM5, LO5, SU4, Be71
	<i>Tubulanus</i> sp.	Ku5, Na5, BM5, LO5, SU5, Be71, LU5
<b>Phoronida</b>	<i>Phoronis</i> sp.	SU5
	<i>Phoronopsis</i> sp.	LO5

**Tab.12:** Determined organisms of the main group Other arranged according to the phylum, and their locations.

## 6.2.5 Polychaeta

Family	Taxa	Stations
<b>Ampharetidae</b>	Ampharetidae <i>Ampharete</i> -like cf. <i>Ampharete</i> sp. <i>Phyllamphicteis</i> ? <i>Phyllamphicteis</i> sp. <i>Phyllocomus</i> sp.	Ku5, Na5, BM5, LO5, SU4, SU5, Be71, LU5 Ku5 BM5, LO5, SU5, Be71 Ku5, Ku4 Na5, LO5, SU4, Be71, LU5 LO5
<b>Amphinomidae</b>	<i>Chloeia</i> sp. <i>Chloeia</i> -like <i>Chloeia inermis</i> Quatrefages, 1866 <i>Paramphinome</i> sp.	LU5 Be71 Be71 Be71
<b>Aphroditidae</b>	Aphroditidae	Na5, SU4, LU5
<b>Capitellidae</b>	Capitellidae Capitellidae? <i>Heteromastus</i> sp. <i>Heteromastus</i> ?	Na5, BM5, SU4, Be71 LO5 Ku4, Be71 Ku5
<b>Chaetopteridae</b>	<i>Chaetopterus</i> sp. <i>Spiochaetopterus costarum</i> (Claparède, 1869)	Ku5, LO5, SU5, Be71, LU5 LO5, LU5
<b>Cirratulidae</b>	<i>Caulleriella</i> sp. <i>Chaetozone</i> B <i>Chaetozone</i> sp. Cirratulidae Cirratulidae (2 small eyes) Cirratulidae B Cirratulidae (orange) Cirratulidae? Cirratulidae D	BM5 SU5 Ku5, BM5, SU5, Be71 Ku5, Ku4, Na5, BM5, LO5, SU4, SU5, Be71, LU5 Ku5 Ku5, BM5 Ku5 LO5 Be71
<b>Cossuridae</b>	<i>Cossura coasta</i> Kitamori, 1960	Ku5, Ku4, Na5, BM5, LO5, SU5, Be71, LU5
<b>Eunicidae</b>	<i>Eunice</i> sp. <i>Marphysa</i> sp.	Be71 Be71
<b>Euphrosinidae</b>	<i>Euphrosine</i> sp.	SU4
<b>Fabriciidae</b>	<i>Fabricia</i> sp.	BM5, SU4, SU5, LU5
<b>Flabelligeridae</b>	Flabelligeridae <i>Pherusa</i> sp.	Na5, LO5 Ku5, Ku4, SU5
<b>Goniadidae</b>	<i>Goniada congoensis</i> Grube, 1877 <i>Goniada</i> sp. Goniadidae	Na5 Ku5, Ku4, SU5, Be71 Na5
<b>Glyceridae</b>	<i>Glycera</i> sp. Glyceridae	Na5 BM5, LO5, SU5, Be71, LU5
<b>Hesionidae</b>	Hesionidae Hesionidae? <i>Ophiodromus</i> sp. <i>Ophiodromus</i> ?	Ku5 Na5 Na5, BM5, LO5, Be71, LU5 BM5
<b>Heterospionidae</b>	<i>Heterospio</i> cf. <i>longissima</i> Ehlers, 1874	SU5

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<b>Heterospionidae</b>	<i>Heterospio</i> sp.	Be71, LU5
<b>Lumbrineridae</b>	Lumbrineridae Lumbrineridae B	Ku5, Na5, BM5, LO5, SU4, SU5, Be71, LU5 BM5
<b>Magelonidae</b>	<i>Magelona</i> sp. Magelonidae	SU4, SU5 BM5, Be71
<b>Maldanidae</b>	<i>Euclymene</i> sp. Maldanidae cf. <i>Petaloproctus</i> <i>Rhodine</i> sp.	BM5 Na5, BM5, LO5, SU4, Be71, LU5 LU5 Ku5, BM5, LO5, SU5, Be71
<b>Nephtyidae</b>	Nephtyidae Nephtyidae?	Ku5, Ku4, Na5, BM5, LO5, SU4, SU5, Be71, LU5 BM5
<b>Nereididae</b>	Nereidae <i>Nereis</i> sp. <i>Pseudonereis</i> B <i>Pseudonereis variegata</i> (Grube, 1857)	Ku5, LU5 Be71 LU5 LO5
<b>Onuphidae</b>	<i>Diopatra</i> juv. <i>Diopatra neapolitana</i> Delle Chiaje, 1841 <i>Diopatra</i> sp. cf. <i>Diopatra</i> cf. <i>Onuphis</i>	Ku4 Ku5, Ku4, BM5, LO5, LU5  Na5, BM5, LO5, SU5, Be71 BM5 BM5
<b>Opheliidae</b>	<i>Ophelia</i> sp.	BM5, SU4
<b>Orbiniidae</b>	<i>Orbinia</i> sp. Orbiniidae Orbinidae? Questidae cf. <i>Questidae</i> ? <i>Scoloplos</i> sp.	BM5 Be71 BM5 SU5 BM5 Na5, BM5, LO5, SU5, Be71
<b>Oweniidae</b>	<i>Myriochele</i> sp. <i>Owenia</i>	Ku5, SU5 Ku4, Na5, BM5, LO5, SU4, SU5, Be71
<b>Paraonidae</b>	<i>Aricidea</i> sp. <i>Aricidea</i> B <i>Aricidea</i> -ähnlich <i>Cirrophorus</i> sp. <i>Cirrophorus</i> B Paraonidae	Na5, BM5, LO5, Be71, LU5 LO5 SU5 Na5, BM5, LO5, Be71 BM5 BM5, SU5, Be71
<b>Pectinariidae</b>	<i>Amphictene</i> sp. <i>Pectinaria</i> sp.	Ku5, SU5, Be71 SU4, Na5, BM5, LO5, LU5, Be71
<b>Phyllodocidae</b>	<i>Eteone</i> sp. <i>Eumida</i> sp. Phyllodocidae	Na5, BM5, SU4, Be71 Ku4, Na5 Na5, LO5, Be71, LU5
<b>Pilargidae</b>	<i>Loandalia</i> sp. <i>Sigambra</i> cf. <i>robusta</i> (Ehlers, 1908) <i>Sigambra</i> sp. <i>Sigambra</i> ?	LO5 Ku5, Ku4 Ku5, Na5, BM5, LO5, SU5, Be71, LU5 LO5
<b>Poecilochaetidae</b>	<i>Poecilochaetus</i> sp. Poecilochaetidae	LO5, Be71 BM5
<b>Polynoidae</b>	<i>Harmothoe</i> sp.	Ku4, Ku5, BM5, LO5, SU5

## 6 Appendix

<b>Polynoidae</b>	<i>Harmothoe</i> B <i>Harmothoe</i> C	LO5 BM5
<b>Sabellariidae</b>	<i>Sabellaria eupomatoides</i> Augener, 1918	Ku4
<b>Sabellidae</b>	<i>Chone</i> sp. <i>Laonome</i> sp. Sabellidae	BM5 LO5, SU5 Na5
<b>Serpulidae</b>	Serpulidae	BM5, SU4, Be71, LU5
<b>Sigalonidae</b>	<i>Sthenelais</i> cf. <i>incisa</i> Grube, 1877 <i>Sthenelais</i> sp.	LO5 BM5, LO5, SU4, SU5, Be71, LU5
<b>Sipunculidae</b>	Sipunculidae	Na5
<b>Spionidae</b>	<i>Laonice</i> B <i>Malacoceros</i> sp. <i>Minuspio</i> ? <i>Paraprionospio pinnata</i> (Ehlers, 1901) <i>Polydora</i> sp. <i>Prionospio</i> B <i>Prionospio sexoculata</i> Augener, 1918 <i>Prionospio</i> sp. <i>Scolelepis</i> sp. <i>Spio</i> -ähnlich <i>Spio</i> B <i>Spiophanes afer</i> Meissner, 2005 <i>Spiophanes</i> black side <i>Spio</i> black side <i>Spio</i> sp. <i>Spiophanes</i> sp.	SU4 SU4 LO5 Ku5, Ku4, BM5, LO5, SU5, Be71, LU5 Na5, BM5, SU4, SU5, Be71, LU5 Ku5, LO5, Be71 LO5, SU4 SU4, Ku5, Ku4, Na5, BM5, LU5, Be71 SU5 LO5 Be71 Be71 Na5, LO5, Be71 BM5, LO5 Ku5, Ku4, Na5, BM5, LO5, Be71, LU5 BM5, SU5, Be71
<b>Sternaspidae</b>	<i>Sternaspis scutata</i> Ranzani, 1817 Sternaspida	LO5, Be71, LU5 Be71
<b>Syllidae</b>	<i>Autolytus</i> sp. <i>Exogone</i> sp. Syllidae <i>Typosyllis</i> sp.	Be71 SU4 Ku5, Na5, BM5, SU4 SU4
<b>Terebellidae</b>	<i>Lanice</i> sp. cf. Terebellidae Terebellidae <i>Terebellides stroemii</i> Sars, 1835	SU5 SU4 Na5, BM5, LO5, Be71, LU5 LO5
<b>Trochochaetidae</b>	<i>Trochochaeta</i> sp.	Na5, LO5, LU5
-	Pointed polychaete Polychaete small Polychaete (brown head) Polychaete small A Polychaete small B Polychaete small C Polychaete small D Scale worm	LO5 Na5 BM5 BM5 BM5 BM5 BM5 Be71

**Tab.13:** Determined polychaetes arranged according to the family, also undetermined polychaetes, and their locations.

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## **Declaration of academic honesty**

I hereby confirm to have written this master's thesis on my own. No other literature sources were used than the listed ones.

Diana Moritz

Rostock, 20<sup>th</sup> August 2012